# TETRASTICHUS GALACTOPUS (HYM.:EULOPHIDAE), A HYPERPARASITE OF APANTELES RUBECULA AND APANTELES GLOMERATUS (HYM.:BRACONIDAE) IN NORTH AMERICA

VINCENT G. NEALIS

Institute of Animal Resource Ecology and Department of Plant Science, University of British Columbia Vancouver, British Columbia 2075 Wesbrook Mall, Vancouver, B.C.

## ABSTRACT

The biology of *Tetrastichus galactopus*, a hyperparasite of *Apanteles rubecula* and *A. glomeratus* in North America, is reviewed. The female hyperparasite attacks its hosts while they are larvae in their primary host, *Pieris rapae*, although free-living cocoons of *A. glomeratus* may also be attacked. In Vancouver, where *A. rubecula* is the only available host, *T. galactopus* is active from July through September and probably overwinters as a pupa or adult within the host cocoon. The significance of hyperparasitism to the success of biological control efforts using *Apanteles* is discussed.

### INTRODUCTION

Tetrastichus glactopus (Ratzburg) is a gregarious eulophid hyperparasite of braconid larvae. In Europe, it has been commonly reared from Apanteles glomeratus (L.) parasitizing Pieris brassicae (L.), the large cabbage white. It is a true hyperparasite, depositing several eggs through the primary host directly into the larval stage of its own host (Picard 1921). Ferrière and Faure (1925) observed oviposition through newly spun cocoons of A. glomeratus but thought a different species of hyperparasite was involved. This view, although not supported by morphological evidence (Delucchi 1950), has contributed to taxonomic confusions; Tetrastichus rapo Walker is the most common synonym in the literature (Krombien et al. 1979). Picard (1921). Gautier and Bonnamour (1924) and Richards (1940) reported hyperparasitism by T. galactopus of Apanteles rubecula Marshall, a solitary parasite of the imported cabbageworm, Pieris rapae (L.). Both A rubecula and A. glomeratus attack early instar P. rapae larvae, emerge from the fourth and fifth instars respectively, and spin a pupal cocoon. The former parasite normally lays one egg to produce a single larva; the latter lays thirty to sixty eggs at a time in each host.

T. galactopus must have entered North America with A. glomeratus which has been widely distributed for biocontrol of P. rapae. In British Columbia, T. galactopus attacks A. rubecula (Wilkinson 1966). Where releases of A. rubecula have been made in areas formerly inhabited by A. glomeratus only, local T. galactopus parasitize the progeny of the released A. rubecula (Parker et al. 1971).

During 1981-82, I made several collections of *T. galactopus* from *A. rubecula* in the lower mainland of British Columbia and established colonies using both *A. rubecula* and *A. glomeratus* stocks. *A. glomeratus* is available in the Okanagan Valley but it has not been collected around Vancouver. It is not

known if *T. galactopus* is also in the Okanagan. This paper reports field and laboratory data and reviews the biology of *T. galactopus*.

# MATERIALS AND METHODS

Field collections made during 1981-82 were from two sites in the Vancouver area. One was a cooperative garden plot allotment near the Fraser River in Burnaby (BBY), which had been one of Wilkinson's (1966) original sites; the other site was an isolated research plot of less than 100 plants, on the Plant Science Field Station at the University of British Columbia (UBC). From both sites in 1981 and at BBY in 1982, P. rapae larvae were brought to the laboratory, separated by instar and reared on potted kale. If Apanteles emerged from the caterpillars, the cocoons were weighed and kept individually in gelatin caps at 22°C and a 16L:8D photoperiod. Apanteles cocoons collected in the field were treated similarly. In 1982 A. rubecula reared at UBC for diapause studies were an additional data source. These primary parasites were allowed to emerge from the host caterpillar and spin a cocoon in the field, then collected and held at 22°C. and a 16L:8D photoperiod. T. galactopus that emerged from these cocoons were sexed and counted. This technique measured the density and sex ratio of T. galactopus per A. rubecula cocoon and permitted estimates of parasitism rates according to the age of the primary host, through the season.

Adult *T. galactopus* live several weeks at low temperatures if water and honey are available. I kept adults in styrofoam cup-cages and exposed *A. rubecula* and *A. glomeratus* to them as cocoons and at various stages within their host larvae. I was thus able to observe the oviposition behaviour of *T. galactopus*. I also exposed non-parasitized cabbageworms to *T. galactopus* as well as final instar *A rubecula* that had experienced a pre-treatment that would induce diapause i.e. less than 14 h photophase. Some *Apanteles* were exposed to un-

**TABLE 1.** Number and per cent parasitism of **Apanteles rubecula** by **Tetrastichus galactopus** according to the stage in which the primary host (**Pieris rapae**) was collected. Data pooled from all collections BBY, 1981 and 1982.

| HOST STAGE COLLECTED | PARASITES          |                             |
|----------------------|--------------------|-----------------------------|
| Pieris rapae         | Apanteles rubecula | Tetrastichus galactopus (%) |
| III                  | 124                | 9 (7.2)                     |
| IV                   | 352                | 132 (37.5)                  |
| A. rubecu/a cocoons  | 94                 | 37 (39.3)                   |

mated female *T. galactopus* to determine the sex of haploid progeny. Attacked larvae were reared individually on plants or, in cases where I intended only to confirm oviposition, were dissected undersaline.

### **RESULTS AND DISCUSSION**

My observations confirm earlier studies that T. galactopus attacks both A. rubecula and A. glomeratus while they are inside the primary cabbageworm host. Although oviposition is normally directly into the primary parasite, a few eggs were found in the caterpillar's body cavity. It is unlikely that these eggs can develop and eventually enter the appropriate host (Richards 1940). Picard (1921) described T. galactopus attacking non-parasitised P. rapae but eggs were never found in the body cavity. I observed T. galactopus inserting their ovipositors through the body wall of several nonparasitised P. rapae but only once were eggs found in the dissected caterpillars. Oviposition must require a further stimulus provided only by the presence of a primary parasite.

Most parasitism of A. rubecula by T. galactopus occurs when the cabbageworm is in the fourth instar, that is, when A. rubecula is close to emergence (Table 1). In laboratory trials, A. rubecula, onequarter and midway through the total parasitic period, were exposed to T. galactopus. The results showed that successful parasitism of A. rubecula was unlikely before the second instar and in most cases not until a few days before emergence. I have never succeeded in obtaining parasitism of A rubecula after the larva has emerged from the primary host and spun a cocoon. T. galactopus walks over A. rubecula cocoons but does not oviposit. The silk may be too dense to permit insertion of the ovipositor. In comparison, T. galactopus parasitizes A. glomeratus either through the primary host or after the free-living cocoon is formed but before pupation of the primary parasite. All individuals in a cluster are not necessarily attacked.

The number of *T. galactopus* emerging from one host cocoon varies widely. More emerge from *A. rubecula* cocoons  $(10.6 \pm 0.89$  hyperparasites per cocoon; range 1-21) than from the smaller cocoons of *A. glomeratus*  $(4.3 \pm 0.52$  hyperparasite per cocoon; range 2-7). The highest hyperparasite densities may be due to superparasitism as more than one *T. galactopus* is often seen on a single primary host in the field. The data relating the density of *T. galactopus* to the weight of *A rubecula* show that host size does not significantly influence parasite density (r = 0.142; df = 52). Rather, a high hyperparasite density per host leads to a smaller average size as has been shown for other gregarious parasites (Bouletreau 1971).

Mature *T. galactopus* larvae bite emergence holes through the host body wall but remain within the host cocoon to pupate. Adults emerge over a protracted period, through one to several holes in the host cocoon. What appears to be a strong positive phototaxis results in rapid dispersal of these adults.

*T. galactopus* is arrhenotokous; unmated females produced only male progency. Most field collected parasitized *A rubecula* yielded both sexes of adult *T. galactopus*; only a small proportion were all female (10.5%) or all male (6.6%). Nevertheless, the sex ratio in the collections after August was always skewed in favour of females and varied through the remainder of the season (Figure 1a). Richards (1940) also found a preponderance of female *T. galactopus* in field samples.

When A. rubecula were reared under field or laboratory conditions that normally induced diapause and were used as hosts of *T. galactopus*, the hyperparasite developed continuously. It is not known if there is a true diapause in *T. galactopus* but if so, it seems from these observations that it is independent of the host's response to environmental conditions.

In Europe, *T. galactopus* is widely distributed over the range of *A. glomeratus*, parasitizing over 50% of the available larvae at some times of the year (Mook and Haeck 1965; Richards 1940). There

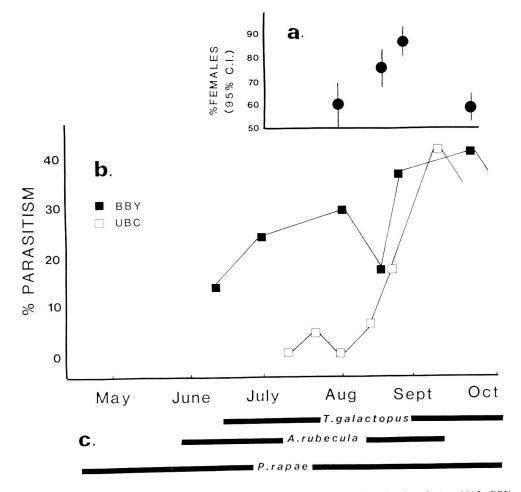


Fig. 1. Phenology of Tetrastichus galactopus at two locations in British Columbia during 1982 (BBY-Burnaby; UBC-University of British Columbia). a) Per cent female T.galactopus at BBY. b) Per cent parasitism of A. rubecula (ex IV-instar P.rapae + cocoons only) by T.galactopus at BBY and UBC. c) Seasonal activity schedule for adult P.rapae, A.rubecula and T.galactopus in the Vancouver area.

are a few estimates for A. rubecula from North America. When A. rubecula was released in Missouri, hyperparasitism by T. galactopus ranged from 36.6% to 72.4% (Parker et al. 1971). At BBY, hyperparasitism of A. rubecula reached 62% in 1981 and 41% in 1982. Comparable levels were observed at UBC in 1982. Except for the relatively late start of the UBC population, the same seasonal pattern was evident at both study sites (Fig 1b). Parasites often lag behind their hosts in spring appearance (compare P. rapae and A. rubecula in Figure 1c). In the case of T. galactopus, the lag may be due to a sustained diapause through the spring or a high winter mortality and perhaps frequent local extinction with subsequent slow recolonization of the sample areas. Mook and Haeck (1965) thought dispersal of T. galactopus was rapid, an impression consistent with T. galactopus finding the isolated cohorts at UBC; but the evidence is circumstantial. The occurence of T. galactopus in the earliest collections at BBY make it arguable that the hyperparasites are highly synchronized with the first generation of the primary parasite and that the apparent time lag is due to previous local extinctions in that area.

It is difficult to ascertain the effects of hyperparasitism on populations of either A. glomeratus or A. rubecula. Blunck (1957) states that hyperparasites in Germany "diminish the useful effect of A. glomeratus" but offers no data. Parker et al. (1971) did not think the "sizeable loss of A. rubecula to hyperparasites throughout the season" affected its role in suppression of P. rapae populations. But it is true that A. rubecula has failed to become established at their release sites in Missouri where hyperparasitism by *T. galactopus* was common. However *A. rubecula's* persistence in Vancouver despite appreciable hyperparasitism by *T. galactopus* argues against hyperparasitism as a predominant limitation to establishing *A. rubecula* in North America.

# **ACKNOWLEDGEMENTS**

All laboratory work was carried out at the Vancouver Research Station, Agriculture Canada. I thank B. D. Frazer for research space and review of the manuscript, and A. T. S. Wilkinson for advice. C. Yoshimoto of the Biosystematics Research Institute, Ottawa confirmed my determination of *Tetrastichus galactopus*.

# REFERENCES

- Blunck, H. 1957. Pieris rapae (L.), its parasites and predators in Canada and the United States. J. Econ. Entomol. 50:835-836.
- Bouletreau, M. 1971. Métabolisme respiratoire de **Pteromalus puparum** (Hym, Chalc.) au cours du développement et influence de la densité de population larvaire. Ann. Zool. Ecol. Anim. 3:195-207.
- Delucchi, V. 1950. Note morfologiche su **Tetrastichus rapo** Walker ,Chalcidide parassita di Imenotteri utili all'agricoltura. Redia 35:441-450.
- Ferrière, C. and J. C. Faure. 1925. Contribution a l'étude des Chalcidiens parasites de l' Apanteles glomeratus, L. Ann. Epiphytes 11:221-234.
- Gautier, C. and S. Bonnamour. 1924. Recherches sur Tetrastichus rapo, (Hym. Chalcididae). Rev. Path. Veg. & Ent. Agric. 11:246-253.
- Krombien, K. B., P. D. Hurd, Jr., D. R. Smith and B. D. Burks. 1979. Catalog of Hymenoptera in America North of Mexico. Vol. I Smithsonian Institution Press, Washington, D.C. 1198pp.
- Mook, J. H. and J. Haeek. 1965. Dispersal of Pieris brassicae L. (Lepidoptera:Pieridae) and of its primary and secondary hymenopterous parasites in a newly reclaimed ploder of the former zuiderzee. Archiv. Neerlandaises de Zoologie 16:293-312.
- Parker, F. D., F. R. Lawson and R. E. Pinnell. 1971. Suppression of Pieris rapae using a new control system: mass releases of both the pest and its parasites. J. Econ. Entomol. 64:721-735.
- Picard, F 1921. Sur la biologie du Tetrastichus rapo Walk. (Hym. Chalcididae). Bull. Soc. Ent. Fr. 9:206-208.
- Richards, O. W. 1940. The biology of the small white butterfly (Pieris rapae), with special reference to factors controlling its abundance. J. Anim. Ecol. 9:243-288.
- Wilkinson, A. T. S. 1966. Apanteles rubecula Marsh and other parasites of Pieris rapae in British Columbia. J. Econ. Entomol. 59:1012-1013.