

MICROORGANISMS ISOLATED FROM FOREST INSECTS OF BRITISH COLUMBIA

O. N. MORRIS¹

Forest Pest Management Institute
Canadian Forestry Service
Sault Ste. Marie, Ontario
P6A 5M7

ABSTRACT

Pathogenic and non-pathogenic microorganisms including fungi, bacteria, viruses, microsporidia and nematodes were isolated from about 14,000 specimens representing 108 pest species of insects collected from British Columbia forests between 1949 and 1969. *Entomophthora* sp. and *Beauveria* sp were the most widely distributed fungal organisms isolated, occurring in 14 and 29 insect species, respectively. Nuclear polyhedrosis and granulosis viruses were isolated from 53 species, microsporidia from 26, pathogenic bacteria from 12 and nematodes from 2 species. A new variety of *Bacillus thuringiensis*, viz. *canadensis*, was isolated from *Lambdina fiscellaria lugubrosa* (Hbst.) and a *Neophasia* sp. The largest numbers of species of microorganisms were found in *Melanolophia imitata* (Wlk.), *Malacosoma disstria* Hbn., *M. pluviale* (Dyar), *L. f. lugubrosa*, *Acleris variana* (Fern.), *Hyphantria cunea* Dru., *Choristoneura fumiferana* (Clem.), *Orgyia pseudotsugata* (McD) and *Neophasia menapia* Feld.

INTRODUCTION

The concept of pest management involves the rational utilization of a variety of biotic and abiotic agents in a well-planned strategy against agricultural and forest insect pests. The naturally occurring microbial control agents can form an important component of such strategies and, indeed, their use presently constitutes a highly advanced alternative to chemical pesticides. However, prior to introducing microbial control agents into a stable forest ecosystem it is essential, for several reasons, that an inventory of the natural enemies of the pest species be taken. Firstly, native biocontrol agents can have devastating effects on the insect populations and their isolation, characterization and manipulation could form part of the management program. Secondly, man-made intervention schemes may not always be compatible with natural biotic schemes already operating in the environment. Finally, the development of simulation models for pest management presupposes a knowledge of the pest's population dynamics which should include key factor analysis of the natural enemy complexes.

An insect disease survey was initiated in 1949 in the forests of British Columbia by the late S. M. Sagar, and continued between 1960 and 1969 by the author. This paper summarizes the results of that survey and, with the possible exception of a similar survey in Quebec (Smirnov and Juneau, 1973), constitutes the most extensive and detailed documentation of microorganisms found in forest insects from a single geographic area in Canada.

MATERIAL AND METHODS

Dead insects, collected by laboratory staff during

the annual insect and disease surveys, were submitted in plastic pill capsules to the Pacific Forest Research Centre and were stored at 4°C until examined for microorganisms. In cases of widespread viral epizootics, large numbers of cadavers were sent collectively to allow for purification of the pathogen for study beyond simple light microscopy identification.

Cadavers with external symptoms of fungal infections were sent directly to the Insect Pathology Research Institute (IPRI), Sault Ste. Marie, Ontario, for identification of the pathogen. Wet mounts of cadavers in which fungal spores, conidia or other development elements were found were isolated on potato-dextrose agar or Sabouraud's medium and sent to IPRI. Occasionally, fungal specimens were identified by the Insect Disease Diagnostic Laboratory, University of California, Berkeley.

Insect specimens were considered positive for virus infection only if virus inclusion bodies were observed in susceptible tissue sites (fat body, hypodermis, tracheal matrix, hemocytes and gut epithelium). In a few cases, when sufficient material was available, nuclear polyhedrosis virus inclusion bodies were purified by differential centrifugation, processed for transmission electron microscopy, and sectioned to reveal the capsid nature of the virus.

Suspected cases of microsporidial infections were either fixed to glass slides and stained with Giemsa to observe the characteristic differential staining of regions of the spore, or pressure was applied to a cover slip above a wet smear to facilitate the extrusion of polar filaments. The identification of protozoan parasites was limited to the Order Microsporidia.

¹Present Address: Agriculture Canada Research Station, 195 Dafoe Road, Winnipeg, Manitoba R3T 2M9.

Most bacteria found in insects are saprophytic aerobic species which may cause lethal septicemia upon entering the hemocoel. Because nearly all the insect species examined during the present survey contained saprophytic or pathogenic bacteria and since time and resources did not allow for identifying all bacterial isolates, only those in which spores, or spores and crystals were present in squashed preparations were identified to species. Eighteen isolates were eventually identified, using reaction to Gram stain, cellular morphology, colony type, slant form, broth form, oxygen requirements, temperature requirements, growth on special media (motility medium, blood agar, NaCl broth) and biochemical reactions. The latter included fermentation reactions in glucose, lactose, maltose, sucrose, xylose, mannitol, arabinose, cellobiose, fructose, galactose, mannose, raffinose, rhamnose,

trehalose, dulcitol, inositol, sorbitol, dextrin, inulin, salicin, starch and levulose. Other biochemical reactions were gelatin liquefaction, indol production, Nitrate reduction, litmus milk, methyl red, H₂S production, V.P. test, urea slant (ammonia), citrate agar, citrate broth, phospholipase C, and casein hydrolysis. On this basis, a preliminary identification of the bacterial species was made, and slant cultures were sent to Dr. H. de Barjac, Institut Pasteur in Paris for species confirmation and serotyping. A total of about 14,000 specimens representing 108 insect species were examined during the 20-year survey periods. The number of distinct locations and the year in which the microorganisms were found are listed in Table 1, indicating geographical distribution and frequency of occurrence. The complexes of microorganisms isolated from the most important economic pests are given in Table 2.

TABLE 1. Microorganisms isolated from forest insects of British Columbia.

Pathogens	Hosts	Number of Locations	Years
FUNGI			
<i>Entomophthora</i> *	<i>Melanolophia imitata</i> (Wlk.)	31	1949, 1950, 1953, 1956, 1957, 1958, 1965, 1967
	<i>Nyctobia limitaria nigroangulata</i> Stkr.	1	1956
	<i>Ectropis crepuscularia</i> Schiff.	1	1950, 1951
	<i>Caripeta divisata</i> (Wlk.)	3	1950
	<i>Lambdina fiscellaria lugubrosa</i> (Hulst.)	6	1947, 1949, 1950, 1951, 1953, 1956, 1967
	<i>Malacosoma pluviale</i> (Dyar)	1	1956
	<i>Nematocampa filamentaria</i> Gn.	1	1954
	<i>Griselda radicana</i> Wlshm.	1	1954
	<i>Acleris variana</i> (Fern.)	2	1956
	<i>Nyctopia phantasmaria</i> (Strk.)	1	1957
	<i>Anthelia hyperborea</i> (Hbst.)	1	1956
	<i>Pikonema dimmockii</i> (Cress.)	1	1954
	<i>Arge pectoralis</i> (Leach)	1	1953
	<i>Neodiprion tsugae</i> (Midd.)	3	1954, 1957
	<i>Beauveria</i> (<i>globilifera</i>) (Speg.)*	<i>Choristoneura fumiferana</i> (Clem.)	5
<i>Malacosoma pluviale</i> (Dyar)		7	1957, 1958
<i>Dendroctonus obesus</i> (Mann)		1	1968
<i>Beauveria</i> <i>bassiana</i> (Bals.)*	<i>Venusia cambrica</i> Cur.	1	1950
	<i>Malacosoma pluviale</i> (Dyar)	7	1957, 1958
	<i>Dendroctonus obesus</i> (Mann)	1	1968
<i>Beauveria</i> sp.*	<i>Lambdina fiscellaria lugubrosa</i> (Hlst.)	2	1953, 1954
	<i>L. f. somniaria</i> (Hlst.)	1	1949
	<i>Epirrita autumnata</i> (Gn.)	1	1949
	<i>Campaea perlata</i> (Gn.)	1	1949
	<i>Enyptia venata</i> (Grt.)	3	1949, 1953
	<i>Enyptia packardata</i> Taylor	1	1953
	<i>Feralia jacosia</i> (Gn.)	1	1958
	<i>Stilpnotia salicis</i> (L.)	1	1956
	<i>Syngraphia alias interalia</i> Ott.	1	1953
	<i>Trichiosoma triangulum</i> Kyb.	2	1951, 1957
	<i>Orgyia pseudotsugata</i> (McD)	1	1958

TABLE 1. (cont'd) Microorganisms isolated from forest insects of British Columbia.

Pathogens	Hosts	Number of Locations	Years
	<i>Archips cerasivorana</i> Fitch	1	1954, 1957
	<i>Epicnaptera americana</i> Harr.	1	1949
	<i>Pseudohazia eglanterina</i> (Bdv.)	1	1957
	<i>Malacosoma disstria</i> Hbn.	1	1954
	<i>Halisidota argentata</i> Pack.	1	1954
	<i>Apantesis parthenia</i> Kby.	1	1954
	<i>Choristoneura fumiferana</i> (Clem.)	2	1957
	<i>Cephalcia</i> sp.	1	1949
	<i>Tolyte</i> sp.	1	1957
	<i>Neodiprion tsugae</i> Midd.	3	1949, 1954
	<i>Hemichroa crocea</i> (Fourc.)	1	1949
	<i>Pityophagus rufipennis</i> Hom.	1	1957
<i>Cephalosporium</i> sp.	<i>Epirrita autumnata</i> (Gn.)	1	1949
	<i>Campaea perlata</i> (Gn.)	1	1949
	<i>Syngrapha alias interalia</i> Ott.	1	1949
	<i>Adelges piceae</i> (Ratz.)	1	1965
	<i>Eupithecia filmata</i> Pears	1	1967
	<i>Pikonema</i> sp.	1	1967
<i>Penicillium</i> sp.	<i>Malacosoma pluviale</i> (Dyar)	2	1953, 1957
	<i>Melanolophia imitata</i> (Wlk.)	2	1951, 1965
	<i>Neodiprion tsugae</i> Midd.	1	1950, 1951
	<i>Adelges piceae</i> (Ratz.)	1	1965
	<i>Lambdina fiscellaria somniaria</i> (Hlst.)	1	1965
	<i>Microlepidoptera</i>	1	1965
<i>Verticillium</i> sp.*	<i>Melanolophia imitata</i> (Wlk.)	1	1949
	<i>Smerinthus cerisyi</i> Kby.	1	1949
<i>Fusarium</i> sp.*	<i>Malacosoma pluviale</i> (Dyar)	1	1956
	<i>Hemichroa crocea</i> (Fourc.)	1	1949
	<i>Neodiprion</i> sp.	1	1949
<i>Alternaria</i> sp.	<i>Malacosoma pluviale</i> (Dyar)	1	1956
	<i>Zale duplicata largera</i> Sm.	1	1957
<i>Spicaria</i> sp.*	<i>Malacosoma pluviale</i> (Dyar)	1	1956
	<i>Acleris variana</i> (Fern.)	1	1957
	<i>Cephalcia</i> sp.	1	1967
<i>Hirsutella</i> sp.*	<i>Nictobia limitaria nigroangularia</i> Strk.	1	1950
<i>Cordyceps</i> sp.*	<i>Melanolophia imitata</i> (Wlk.)	1	1961
<i>Aspergillus</i> sp.	<i>Malacosoma pluviale</i> (Dyar)	1	1965
<i>Harmodendrum</i> sp.	<i>Acleris variana</i> (Fern)	1	1967
	<i>Melanolophia imitata</i> (Wlk.)	1	1967
<i>Trichosporon</i> sp.	<i>Melanolophia imitata</i> (Wlk.)	1	1968
VIRUS			
<i>Baculovirus</i>	<i>Lambdina fiscellaria lugubrosa</i> (Hlst.)	17	1947, 1949, 1951, 1953, 1954, 1956
(Nuclear Polyhedrosis virus subgroup) (NPV)	<i>Nepytia phantasmaria</i> (Stkr.)	3	1956, 1957
	<i>Melanolophia imitata</i> (Wlk.)	10	1950, 1951, 1957, 1958, 1960, 1961, 1965, 1969
	<i>Malacosoma disstria</i> Hbn.	22	1951, 1953, 1954, 1957, 1962, 1968, 1969
	<i>Lambdina fiscellaria somniaria</i> (Hlst.)	1	1948, 1949, 1951, 1958, 1960, 1961, 1962
	<i>Choristoneura fumiferana</i> (Clem.)	4	1950, 1968, 1969
	<i>Ectropis crepuscularia</i> Schiff.	2	1960, 1961, 1962, 1965, 1969
	<i>Malacosoma pluviale</i> (Dyar)	17	1953, 1956, 1957, 1958,

TABLE 1. (cont'd) Microorganisms isolated from forest insects of British Columbia.

Pathogens	Hosts	Number of Locations	Years
			1961, 1962, 1963, 1965, 1966, 1967, 1968, 1969
	<i>Enypia venata</i> Grt.	2	1949
	<i>Hydriomena nubilofasciata</i> Pack.	1	1950
	<i>Hydriomena irata</i> Sw.	1	1950
	<i>Pero mizon</i> Rindge	1	1953
	<i>Pero behrensarius</i> Pack.	1	1956
	<i>Euthyatira pudens</i> Gn.	1	1949
	<i>Feralia jacosa</i> Gn.	2	1949, 1950
	<i>Orthosia hibisci</i> Gn.	2	1949, 1950
	<i>Stilpnotia salicis</i> L.	5	1956, 1958, 1962, 1965, 1969
	<i>Synaxis pallulata</i> (Hlst.)	2	1951, 1954
	<i>Synaxis jubararia</i> (Hlst.)	1	1956
	<i>Syngrapha selecta</i> (Wlk.)	1	1957
	<i>Nyctobia limitaria nigroangulata</i> Stkr.	2	1950, 1965
	<i>Erranis vancouverensis</i> (Hlst.)	2	1958
	<i>Orgyia pseudotsugata</i> (McD.)	8	1954, 1958, 1961, 1962, 1969
	<i>Nymphalis antiopa</i> L.	2	1956, 1968
	<i>Caripeta divisata</i> (Wlk.)	3	1952, 1961
	<i>Hesperumia sulphuraria</i> Pack	1	1949
	<i>Protoarmia indicataria</i> (Wlk.)	1	1949
	<i>Hyperetis amicaria</i> H & S	1	1957
	<i>Dyoryctria pseudotsugella</i> Monroe	2	1962, 1966
	<i>Halisidota argentata</i> Pack	3	1961, 1968
	<i>Acleris variana</i> (Fern.)	25	1954, 1956, 1957, 1960, 1965, 1966, 1967, 1968
	<i>Panthea portlandia</i> Grt.	1	1953
	<i>Orgyia antiqua badia</i> (Hy. Ed.)	2	1954, 1956
	<i>Polygonia satyrus</i> Ed.	1	1956
	<i>Operophtera bruceata</i> (Hlst.)	2	1957
	<i>Eupithecia</i> sp.	3	1956, 1965
	<i>Neodiprion tsugae</i> Midd.	5	1950, 1956, 1957
	<i>Neodiprion abietis</i> (Harr.)	1	1957
	<i>Hemichroa crocea</i> (Fourc.)	1	1950
	<i>Neodiprion</i> sp.	38	1950, 1954, 1956, 1957, 1965
	<i>Eupithecia annulata</i> (Hlst.)	1	1965
	<i>Zeiraphera pseudotsugana</i> Kft.	1	1965
	<i>Pikonema dimmockii</i> (Cress.)	1	1965
	<i>Vanessa cardui</i> Dyar	1	1966
	<i>Neophasia menapia</i> Feld	2	1968
	<i>Tetropium cunnamopterum</i> Kirby	1	1968
	<i>Papilio daunis</i> Bdv.	1	1968
	<i>Hyphantria cunea</i> (Drury)	1	1969
(Granulosis virus subgroup) (G.V.)	<i>Lambdina fiscellaria lugubrosa</i> (Hlst.)	4	1947, 1953
	<i>Lambdina fiscellaria somniaria</i> (Hlst.)	1	1948
	<i>Malacosoma pluviale</i> (Dyar)	3	1953, 1956
	<i>Acleris variana</i> (Fern.)	2	1953, 1954
	<i>Pseudohazia eglanterina</i> (Bdv.)	1	1953
	<i>Griselda radicana</i> Wlsh.	1	1954
	<i>Nematocampa filamentaria</i> Gn.	1	1954
	<i>Sciaphila duplex</i> Wlsh.	1	1957
	<i>Clepsis persicana</i> (Fitch.)	1	1957
	<i>Hyphantria cunea</i> (Drury)	2	1949, 1969
	<i>Arge pectoralis</i> (Leach)	1	1954

TABLE 1. (cont'd) Microorganisms isolated from forest insects of British Columbia.

Pathogens	Hosts	Number of Locations	Years
PROTOZOA			
Microsporidia	<i>Choristoneura fumiferana</i>	6	1950, 1954, 1968, 1969
	<i>Choristoneura conflictana</i>	1	1957
	<i>Acleris variana</i> (Fern.)	8	1953, 1956, 1966, 1968
	<i>Sciaphila duplex</i> Wlshm.	1	1957
	<i>Malacosoma pluviale</i> Dyar	3	1965, 1967, 1969
	<i>Malacosoma disstria</i> Hbn.	2	1968, 1969
	<i>Melanolopia imitata</i> Dyar	2	1965, 1969
	<i>Eupithecia filmata</i> Pears	1	1965
	<i>Orthosia hibisci</i> Gn.	1	1965
	<i>Nyctobia limitaria nigroangulate</i> Stkr.	4	1965, 1966
	<i>Lambdina fiscellaria lugubrosa</i> Hulst.	2	1965, 1966
	<i>Neophasia menapia</i> (Feld.)	1	1965
	<i>Syngrapha</i> sp.	1	1965
	<i>Lithophane lepida</i> (Lintner)	2	1966, 1968
	<i>Pristiphora erichsonii</i> (Hartwig)	3	1966
	<i>Zeiraphera</i> sp.	3	1966, 1969
	<i>Xylotype</i> sp.	1	1966
	<i>Eupisilia</i> sp.	1	1966
	<i>Dendroctonus obesus</i> (Man.)	1	1968
	<i>Zeiraphera destitutana</i> (Walker)	1	1968
	<i>Halisidota argentata</i> Pack	1	1968
<i>Orgyia pseudotsugata</i> Midd.	1	1969	
<i>Leucobrephephos brephoides</i> (Wlk.)	1	1969	
<i>Pikonema dimmockii</i> (Cress.)	2	1968	
BACTERIA			
<i>Bacillus thuringiensis</i> var. <i>galleriae</i>	<i>Acronicta grisea</i> (Wlk.)	1	1965
	<i>Neophasia menapia</i> (Feld.)	1	1965
	<i>Vanessa cardui</i> L.	1	1966
	<i>Malacosoma pluviale</i> Dyar	1	1966, 1967
	<i>Pristiphora erichsonii</i> (Hartig)	1	1966
	<i>Neodiprion</i> sp.	1	1966
<i>Bacillus thuringiensis</i> var. <i>thuringiensis</i>	<i>Pristiphora erichsonii</i> (Hartig)	1	1965, 1966
<i>Bacillus cereus</i>	<i>Eupithecia annulata</i> (Hlst.)	2	1965
	<i>Lambdina fiscellaria lugubrosa</i> Hulst.	1	1965
	<i>Orthosia hibisci</i> Gn.	1	1965
	<i>Melanolophia imitata</i> (Wlk.)	1	1965
<i>Bacillus brevis</i>	<i>Melanolophia imitata</i> (Wlk.)	1	1965
<i>Bacillus thuringiensis</i> var. <i>canadensis</i>	<i>Lambdina fiscellaria lugubrosa</i> (Hlst.)	1	1966
	<i>Neophasia</i> sp.	1	1966
NEMATODES			
Unidentified	<i>Dendroctonus engelmanni</i> Hopk.	1	1950
	<i>Trypodendron linneatum</i> (Oliv.)	1	1962

*Indicates pathogenic fungi

RESULTS AND DISCUSSION**Fungal Pathogens**

Entomophthora and *Beauveria* were the most frequently occurring, and the most widely distributed, fungal microorganisms recorded among forest insects of British Columbia (Table 1).

Entomophthora affected 14 species and were found most frequently in two lepidopterous species, viz. *Melanolophia imitata* (Wlk.), and *Lambdina fiscellaria lugubrosa* (Hlst.). *Entomophthora* spp. have been reported in numerous other forest insects species in all provinces of Canada (MacLeod 1956;

TABLE 2. Common forest insects of British Columbia and microorganisms found in them.

Host	Fungus	Virus	Bacteria	Protozoa	Nematode
<i>Melanolophia imitata</i>	<i>Entomophthora</i> sp. <i>Penicillium</i> sp. <i>Verticillium</i> sp. <i>Cordyceps</i> sp. <i>Harmodendrum</i> sp. <i>Trichosporon</i> sp.	NPV	<i>Bacillus cereus</i> <i>Bacillus brevis</i>	Microsporidia	—
<i>Malacosoma disstria</i>	<i>Entomophthora</i> sp. <i>Beauveria</i> sp.	NPV	—	Microsporidia	—
<i>Malacosma pluviale</i>	<i>Entomophthora</i> sp. <i>Beauveria bassiana</i> <i>Penicillium</i> sp. <i>Fusarium</i> sp. <i>Alternaria</i> sp. <i>Spicaria</i> sp. (<i>Noumeria</i>) <i>Aspergillus</i> sp.	NPV GV	<i>Bracillus thuringiensis</i> var. <i>galleriae</i>	Microsporidia	—
<i>Ectropis crepuscularia</i>	<i>Entomophthora</i> sp.	NPV	—	—	—
<i>Caripeta divisata</i>	<i>Entomophthora</i> sp.	NPV	—	—	—
<i>Lambdina fiscellaria lugubrosa</i>	<i>Entomophthora</i> sp. <i>Beauveria</i> sp.	NPV GV	<i>Bacillus cereus</i>	Microsporidia	—
<i>Lambdina fiscellaria somnaria</i>	<i>Beauveria</i> sp. <i>Penicillium</i> sp.	NPV GV	—	—	—
<i>Acleris variana</i>	<i>Entomophthora</i> sp. <i>Spicaria</i> sp.	NPV GV	—	Microsporidia —	— —
<i>Neodiprion tsugae</i>	<i>Entomophthora</i> sp. <i>Beauveria</i> sp. <i>Penicillium</i> sp. <i>Fusarium</i> sp.	NPV	—	—	—
<i>Choristoneura fumiferana</i>	<i>Entomophthora</i> sp. <i>Beauveria globulifera</i>	NPV	—	Microsporidia	—
<i>Hyphantria cunea</i>	<i>Beauveria globulifera</i>	NPV GV	—	—	—
<i>Stilpnotia salicis</i>	<i>Beauveria</i> sp.	NPV	—	—	—
<i>Orgyia pseudotsugata</i>	<i>Beauveria</i> sp.	NPV	—	Microsporidia	—
<i>Adelges piceae</i>	<i>Cephalosporium</i> sp. <i>Penicillium</i> sp.	—	—	—	—
<i>Pristiphora erichsonii</i>	—	—	<i>Bacillus thuringiensis</i> var. <i>galleriae</i>	Microsporidia	—
<i>Nepytia phantasmaria</i>	—	NPV	—	—	—
<i>Nyctobia limitaria nigroangulata</i>	<i>Hirsutella</i> sp.	NPV	—	Microsporidia	—
<i>Halisidota argentata</i>	—	NPV	—	Microsporidia	—

TABLE 2. (cont'd)

<i>Orygia antiqua badia</i>	—	NPV	—	—	—
<i>Neophasia annulata</i>		NPV <i>Bacillus</i>		Microsporidia	—
		<i>thuringiensis</i>			
		var. <i>galleriae</i>			
<i>Eupithecia annulata</i>		NPV <i>Bacillus cereus</i>		—	—
<i>Choristoneura conflictana</i>	—	—	—	Microsporidia	—
<i>Dendroctonus engelmanni</i>	—	—	—	—	Nematode
<i>Trypodendron lineatum</i>	—	—	—	—	Nematode

— Indicates microorganisms not found.

MacLeod and Muller-Kogler 1973; Smirnof and Juneau 1973; Burke 1980) and have a world-wide distribution.

Beauveria spp. were isolated from 29 insect species in widespread localities of the province but, generally, the frequency of occurrence was low. Smirnof and Juneau (1973) reported that 28 insect species were infected by this pathogen in Quebec. *B. globilifera* isolated from three species, is often treated as a strain of *B. bassiana* (MacLeod 1954). *Cephalosporium* sp. were isolated from 6 insect species but the frequency of occurrence was low and geographical distribution was limited throughout the survey period. This fungus is considered by some taxonomists to be only a varietal form of *Verticillium* sp. (Samson 1981) and has been isolated in Quebec from *Toumeyella numismaticum* CP. AND M. (Smirnof and Juneau 1973). Nine other distinct genera of fungi, viz. *Penicillium*, *Fusarium*, *Alternaria*, *Spicaria*, *Hirsutella*, *Cordyceps*, *Aspergillus*, *Harmodendrum* and *Trichosporon* were isolated from various pest species but with low frequency and restricted distribution.

Viruses

Nuclear polyhedrosis viruses were isolated from 48 host species of insects, occurring most frequently and with the widest geographical distribution among *L. f. lugubrosa*, *M. imitata*, *Malacosoma disstria* Hbn., *L. f. somnaria* (Hlst.), *Malacosoma pluviale* (Dyar), *Ectropis crepuscularia* Schiff., *Stilpnotia salicis* L., *Orygia pseudotsugata* (McD.), *Acleris variana* (Fern.) and *Neodiprion* sp. Electron microscopy of all the Lepidoptera inclusion bodies indicated the presence of multicapsid type of virus (Morris and Olsen 1970). Studies on the histopathology, infectivity and epizootiology of some of these viruses have been reported elsewhere (Morris 1962a, 1962b; 1963a, 1963b; 1967). Granulosis viruses were isolated from 11 lepidopterous species but their frequency and distribution were generally low. Nuclear polyhedrosis and granulosis viruses have been isolated from 31 forest insect species in Quebec (Smirnof and Juneau 1973).

Microsporidia

Microsporidia were identified in 26 insect species but their incidence and geographic distribution among individual insect species were generally low. The exceptions included Microsporidia recorded in *Choristoneura fumiferana*, *Acleris variana*,

Malacosoma pluviale and *Nyctobia limitaria nigroangulata*. Smirnof and Juneau (1973) reported that microsporidia were found in 48 species of forest insects in Quebec, including *C. fumiferana*, *C. conflictana*, *M. disstria* and *Pristiphora erichsonii*. Wilson and Burke (1971) and Wilson (1975) have described the microsporidia from *C. fumiferana* and *C. conflictana*. Thomson's (1960) list of 131 valid species of microsporidia from a vast array of insect species indicates the wide occurrence of these pathogenic microorganisms.

Bacteria

Bacillus spp. were isolated from 56 insect species but only 12 isolates were fully identified. These isolates included *B. thuringiensis* var. *galleriae* from 6 insect species, *B. thuringiensis* var. *thuringiensis* from 1, *B. cereus* from 4, *B. brevis* from 1 and *B. thuringiensis* var. *canadensis* from 2. The bacterial variety, *canadensis*, isolated from *L. f. lugubrosa*, the western hemlock looper, and from a *Neophasia* sp. from Vernon, B.C., is internationally recognized as a type variety because of its stable crystal-forming characteristic. This variety has also been isolated from *Diaropsis* sp. and *Anisacta* sp. in Chad and from *Melolontha* sp. in Madagascar (de Barjac and Bonnefoi 1972). Smirnof and Juneau (1973) reported *B. cereus* from 21 species, *B. thuringiensis* var. *thuringiensis* from 1 and *Bacillus* sp. from 14 forest insect species in Quebec.

Little is known about the epizootiology of bacterial pathogens of forest insects and the impact of these microorganisms on the population dynamics of pest species is relatively obscure. As a group, they probably have little impact compared with that of the viruses, fungi and microsporidia because reported natural epizootics are rare.

Nematodes

Nematodes were isolated from 2 coleopterous species from the Bolean and Cowichan Lake areas. The impact of nematode infections on forest insect populations is unclear but recent work in agriculture seems to justify continued study of these pathogens for use in integrated control programs. (Finney 1981).

Complex of Microorganisms Isolated from Common Forest Insects

The list of common forest insect pests of B.C., with their corresponding microbial isolates (Table 2), shows that some pest species are subject to considerably greater pressure from natural

microorganisms than others. The green-striped forest looper, *M. imitata* for example, occurs in western Alberta and throughout British Columbia as a periodic defoliator of western hemlock, *Tsuga heterophylla* (Raf.) Sarg. and Douglas fir, *Pseudotsuga menziesii* (Mirb.) Franco. It is attacked by 3 genera of fungi viz. *Entomophthora* sp., *Verticillium* sp. and *Cordyceps* sp., a nuclear polyhedrosis virus, a microsporidian parasite and 2 species of pathogenic bacteria. The tent caterpillars, *M. disstria* and *M. pluviale* are also attacked by 3 pathogenic fungi, viz. *Entomophthora* sp., *B. bassiana* and *Spicaria* sp., 2 viruses, a microspori-

dian and a bacterium. Other important pest species which appear to be highly subject to natural diseases include the western hemlock looper, *L. f. lugubrosa*, the black-headed budworm, *A. variana*, the western spruce budworm, *C. fumiferana*, the fall webworm, *Hyphantria cunea*, the douglas fir tussock moth, *O. pseudotsugata* and the pine butterfly, *N. menapia*. It is likely that microorganisms play a role in the dynamics of some important economic forest pests of British Columbia and research into their manipulation and their use in pest management strategies seems justifiable.

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